ZIPGRAM

RESHWATER MUSSEL (1)

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ABSTRACT Ligumia subrostrata (Say) will maintain a steady state for Na when acclimated to an artificial pondwater. Prostaglandin E2 (0.6 $\mu\text{M/g}$ dry tissue) injected into the blood cause an 80% reduction of Na influx and the animals experience a net loss of Na. Injections of indomethacin (0.1 $\mu\text{M/g}$ dry tissue), a prostaglandin synthetase inhibitor, cause a doubling of Na influx relative to control animals. Prostaglandin-like material can be extracted from the blood of L. subrostrata and may be a component of the endocrine control of ion regulation in freshwater bivalves.

Freshwater bivalves maintain their body fluids hyperosmotic to the environment with Na being the principal cation. Blood Na concentration ranges between 15-25 mM/l in the various freshwater mussels which have been studied (Dietz, '77, '79). Recently, we observed that freshwater mussels alsolay a diurnal rhythm in blood Na concentration (Yeider and Dietz, '78). The noted that the changes in blood ion concentration correlated with changes in rates of Na transport suggesting an endogenous control mechanism. In addition, a transient stimulation of Na transport has been observed in the massel Margaritifera hembeli in response to handling (Dietz, '79). These studies suggest a hormonal control mechanism is functioning in Na regulation. Since prostaglandins have been reported in marine bivalvés (Freas, '78) we have examined the effects of prostaglandins on Na transport in freshwater suggests. In this report, we present evidence that prostaglandins are capable

of inhibiting Na transport in the freshwater unionid <u>Ligumia</u> <u>subrostrati</u> MATERIALS AND METHODS <u>Ligumia subrostrata</u> were obtained from local ponds and acclimated to an artificial pondwater (0.5 or NaCl, 0.4 CaCl $_{2\times}$ 0.2 NaHCO $_3$, 0.05 KC1 in mM/1). The mussels were rinsed for 30 min. in water before injection of the drugs. All drugs were injected into the foot tissue, and the animals were returned to distilled water for equilibrium Those animals that opened within I hour after the injection were used to a second seco transport studies. For the flux studies, each animal was placed in a transport studies. container of 0.5 mM 22 Na $_2$ SO $_4$. Bath samples were taken on an hourly begin Sodium concentrations were determined by flame photometry and radioactive determined with a liquid scintillation counter (Triton-X 100, p-terpher, toluene counting fluid). The animals were shucked, dried 90-100 $^{\circ}$ C and with The drugs used were indomethacin (Sigma), arachidonic acid (Sigma), and $p_{\rm col}$ glandin E₂ (Upjohn).

The net flux (J_n) for sodium was determined by the changes in the social concentration of the bath. The unidirectional influx (J $_{\rm j}$) was ${\rm determines}$ the disappearance of $^{22}\mathrm{Na}$ from the bathing media (Dietz and Branton, the efflux (J_0) was estimated by the difference:

$$J_n = J_i - J_o.$$

Blood was collected from animals by pericardial puncture (Fyhn and Costlow, '75). Samples were centrifuged at 8000 X g for 10 minutes at 2-7 to prepare them for prostaglandin extraction (Unger et al., '71). The results extract was separated and visualized on chromatograms (Woods and ${\sf Jocoy},$ The standards used to identify the unknown extraction products of the blood were arachidonic acid, PGE_2 , PGE_1 , PGF_{2a} (Upjohn).

The accuracy of the extraction procedure was checked by the percent recovery of tritium in blood samples spiked with $^3 ext{H-arachidonic}$ acid (Among the effect of dr injection * in tissue; ETOH - J.02; 29 _{leter}thacin - - 0.01; l PGE 2 -) s 0.04; 5 ... ridonic Aci introls for and were poo * <0.05 Pad.01 ing extract of Approximately 4 na vidual coun

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TABLE 1 $$_{\odot}$$ effect of drug treatments on the flux rates of sodium from 0.5 mM $\rm NA_2SO_4$

injection Mary tissue; N)	$\mu Eq/g$ dry tissue-hr $(\overline{X} \pm SE)$			
	Γ	1	U	
ETOH ^a 2 ± 0.02; 29)	0.32 ± 0.25	1.53 ± 0.17	1.21 ± 0.18	
**************************************	2.49 ± 0.32**	3.34 ± 0.15**	0.85 ± 0.18	
PGE ₂ 163 ± 0.04; 5)	-1.16 ± 0.73*	0.29 ± 0.17**	1.45 ± 0.72	
emidonic Acid 198 ± 0.41; 15)	-1.34 ± 0.37**	1.45 ± 0.31	2.79 ± 0.53**	

igntrols for the different treatments were not significantly different and were pooled.

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extract of spiked blood was separated and visualized on a chromatogram. Similarly 4 mm sections of the 80 mm chromatogram strip were scraped into dual counting vials and counted to determine the radioactive R_f values. Differences between groups were determined using the student "t" test were considered significant if P<0.05.

RESULTS Injections of indomethacin caused a significant stimulation ment flux of sodium compared to control animals injected with either modern distilled water vehicles (fig. 1). The indomethacin treated animals decienced a net uptake of Na from the pondwater bath for at least 9 hrs.

The the injection. The control animals (DW and ETOH) were not significantly deferent and remained essentially in a steady state. All of the animals remained in a Cl steady state.

The net uptake of Na in the indomethacin injected animals is due to a

significantly increased influx of Na with no change in the efflux when to ethanol injected controls (table 1). Injections of PGE_2 and its precarachidonic acid, caused depressions of the net flux when compared to the ethanol controls. However, the losses of sodium apparently involved different mechanisms. PGE_2 injections caused a significant decrease of influx rate with the efflux not changed from the controls. Arachidonic different caused the loss of sodium by significantly increasing the efflux with the influx remaining constant.

To determine if these freshwater mussels have endogenous prostagioned like material, we extracted blood from animals acclimated to pondwater blood extracts were chromatographically identified as arachidonic acid property and PGE2 (Rf 28). We frequently observed substantial spots at Rf 38 and 4 which may have been keto derivatives of prostaglandins. When arachidonic was added to control blood samples, the recovery of tritium labeled arachidonic acid was 86%.

DISCUSSION Sodium transport in freshwater mussels is apparently required in part, by prostaglandins. Indomethacin is an effective inhibitor of proglandin synthetase and would prevent the endogenous synthesis of prostaglar from cellular aracidonic acid (Hansen, '74). At the dosage of indomethating injected (about 0.1 μ M/10 g wet tissue), the animals experience a positive balance for several hours. The effect of indomethacin is primarily a structure of the influx of Na. The action of indomethacin is specific for

FIGURE LEGENDS

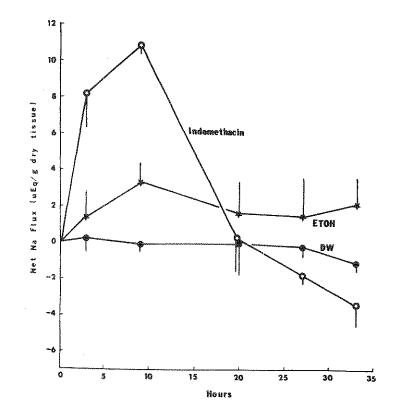
¹ The effect of injections of indomethacin (0.15 \pm 0.02 $\mu\text{M/g}$ dry tiss = 95%ETOH (10 $\mu\text{I/animal}$), distilled water (10 $\mu\text{I/animal}$) on net flux stack point represents 4 animals and the vertical line is 1 SEM.

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Na transport since the animals simultaneously remained in a CI steady state

When the animals were injected with prostaglandin E_2 , they experiences a net loss of Na. The PGE_2 significantly depressed the influx of Na with reffect on the efflux. These data suggest PGE_2 interacts with the epitheliar cells responsible for Na accumulation from the bath. The drug apparently to no effect on the renal tissue since the urinary Na loss was unchanged.

Arachidonic acid, a precursor of prostaglandins, apparently acts intrace by increasing the efflux. These data suggest the arachidonic acid primarial affects the renal tissue at the dose injected. Because of the rapid metators of arachidonic acid in bivalves, there may not have been sufficient PGE synthesis in pondwater acclimated mussels to inhibit the Na influx. However in preliminary studies using salt depleted mussels, we have observed arachident acid to cause a significant reduction of Na influx.

Sodium transport in freshwater bivalves is controlled by endogenous recommendations. Previous reports demonstrated that salt depletion leads to enhanced Na influx (Dietz and Branton, '75; Dietz, '78). The data reported here suggest that prostaglandins are responsible for suppressing epithelial Na influx. It addition, there may be separate controls for the renal tissue. Prostaglanding have been noted to influence Na transport in a variety of tissues (Zins, '75). Declusin et al., '74; Lee, '74). The synthesis of prostaglandins has been reported in marine bivalves (Freas, '78) and we have extracted prostaglanding like material from the blood of freshwater mussels. However, this is the first report of prostaglandin control of Na transport in freshwater bivalves.

ACKNOWLEDGMENTS We thank Ms. D. McMullan for typing the manuscript and Dr. J. E. Pike for providing the prostaglandins.

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Supported by NSF BMS75-05483 A01.

This work was done in partial fulfillment of the requirement for the degree of master of science.

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